Patchy and Janus Nanoparticles by Self-Organization of Mixtures of Fluorinated and Hydrogenated Alkanethiolates on the Surface of a Gold Core

Maria Šologan,1 Domenico Marson,‡ Stefano Polizzi,§ Paolo Pengo,§ Silvia Boccardo,‡ Sabrina Pricl,‡ Paola Posocco,‡,† and Lucia Pasquato*†,‡,⊥

†Department of Chemical and Pharmaceutical Sciences and INSTM Trieste Unit, via L. Giorgieri 1, and ‡MOSE Laboratory, Department of Engineering and Architecture and INSTM Trieste Unit, University of Trieste, Piazzale Europa 1, 34127 Trieste, Italy
§Department of Molecular Sciences and Nanosystems, University Ca’ Foscari of Venezia, via Torino 155/b, I-30172 Venezia-Mestre, Italy

Supporting Information

ABSTRACT: The spontaneous self-organization of dissimilar ligands on the surface of metal nanoparticles is a very appealing approach to obtain anisotropic “spherical” systems. In addition to differences in ligand length and end groups, a further thermodynamic driving force to control the self-assembled monolayer organization may become available if the ligands are inherently immiscible, as is the case of hydrogenated (H-) and fluorinated (F-) species. Here, we validate the viability of this approach by combining 19F NMR experiments and multiscale molecular simulations on large sets of mixed-monolayer-protected gold nanoparticles (NPs). The phase segregation of blends of F- and H-thiolates grafted on the surface of gold NPs allows a straightforward approach to patterned mixed monolayers, with the shapes of the monolayer domains being encoded in the structure of the F/H-thiolate ligands. The results obtained from this comprehensive study offer molecular design rules to achieve a precise control of inorganic nanoparticles protected by specifically patterned monolayers.

KEYWORDS: anisotropic nanoparticles, phase segregation, self-assembly, gold nanoparticles, multiscale molecular modeling, NMR

Patchy particles display precisely controlled surface areas with different properties. These different surface areas determine particle–environment interactions, as nicely demonstrated by several examples of naturally occurring anisotropic particles such as pollen grains1 or viruses.2 Surface morphology is also responsible for the biological activity of these systems; thus, for instance, Janus-like structures of hydrophobin proteins can have roles in the attachment of fungi to different surfaces.3 The development of artificial (nano)particles with anisotropy in shape, surface properties, and chemical functionality will certainly lead to innovative materials.4–7 Indeed, taking inspiration from Nature, chemists now utilize patchy particles as elemental building blocks for the generation of new materials, mainly by exploiting their self-assembly properties in the design of 2D or 3D networks. By this approach, hierarchical structures with applications in biology/medicine and in the mimicry of viruses have been obtained.8–11 This would have been difficult or even out of reach using colloids displaying solely isotropic interactions.12

While many studies carried out in this field concern polymeric particles with size greater than 100 nm, hybrid organic–inorganic nanoparticles characterized by dimensions less than 100 nm have been much less explored.

In this context, the preparation of spherical metal nanoparticles decorated by anisotropic organic monolayers remains challenging, notwithstanding the easy and precise control of their size, surface functionalization, and dispersion. Also, because of their inorganic core, these systems may be endowed with interesting additional properties with respect to soft (e.g., polymeric) particles. In this direction, tremendous efforts have been devoted to theoretical and experimental studies aimed at inducing the organization of thiolate ligands on the surface of gold nanoparticles (NPs) into well-defined domains as a source of anisotropy. Indeed, several different approaches for the

Received: June 14, 2016
Accepted: September 23, 2016
Published: September 23, 2016

DOI: 10.1021/acsnano.6b03931
ACS Nano 2016, 10, 9316—9325
preparation of patchy or Janus hybrid organic—inorganic NPs have been reported. Particularly attractive in this respect is the formation of anisotropic spherical nanoparticles by exploiting the spontaneous self-assembly of coded ligands on their surface. This strategy was pioneered by the group of Stellacci, using mixtures of immiscible thiolate ligands that self-sort on the gold NP surface forming stripe-like domains. These experimental results were coupled to computational investigations carried out by Glotzer and co-workers, who identified the difference in ligand length and the NP core size as two critical parameters in determining the morphology of self-assembled monolayers (SAMs). Thus, according to these seminal studies, binary mixtures of thermodynamically incompatible thiolates can originate three main SAM morphologies on a gold NP surface, depending on their structure and on NP core size: Janus NPs are foreseen when the ligands have comparable length or the NP core diameter is small (typically below 2.5 nm); patches or stripe-like domains are predicted when the two ligands differ in size, while a random organization of the monolayer is expected when one of the two ligands has a branched structure. According to this rationale, several thiol combinations, such as mercaptopropionic acid and octane- or dodecanethiol, mercaptobanesulfonate and phenylmethanethiol, thiopropin and mercaptoundecyltetraethylene glycol, 11-mercapto-1-undecanesulfonate and octanethiol, octanethiol and N-1-[2-[2-(2-methoxyethoxy)ethoxy]ethyl]-8-sulfanylcananamide (HS-C8-TEG), and dodecanethiol and diphenyl thiol, have been experimentally explored.

In this respect, we reasoned that combining H- and F-alkanethiols in the NP decorating monolayer should allow the ligand immiscibility (Flory–Huggins $\chi$ parameter) to be maximized and self-sorting to be promoted, in analogy to a 2D SAM.

In addition, the presence of fluorinated ligands in the coating of hybrid organic—inorganic nanoparticles is of particular interest in material science because of their electronic properties, which may influence the features of the metal core. Moreover, their antifouling surface properties may be exploited at the interface with complex fluids or, even more importantly, in their interactions with biological tissues. So far, these features are unexplored for monolayer-protected metal nanoparticles due to limited examples of fluorinated nanoparticles that are soluble in organic or aqueous solvents.

In this context, we reported examples of 3D mixed monolayers composed of F- and H-thiolates ending with a polyethylene glycol (PEG) group and demonstrated that phase segregation occurs at all molar fractions explored; moreover, we found that the difference in ligand chain length determines the formation of ribbon-like domains on nanoparticles with a gold core larger than 2 nm. Yet, the presence of a PEG terminal group did not allow for a complete dissection of the contribution of the immiscible hydrocarbon/fluorocarbon chains to the morphology of the monolayer.

To unravel this contribution and to further analyze the ligand structural requirements needed to induce well-defined morphologies, we embarked in the synthesis and systematic morphological investigation of gold nanoparticles protected by blends of F- and H-alkanethiols differing in chain length, steric bulk, and relative ratio. This study also allows assessing how the morphology correlates with NP properties such as solubility or aggregation.

Thus, we prepared 2–4 nm core diameter gold NPs decorated by blends (Figure 1) of F- and H-ligands of (i) different length: NP-C16/F6, protected by blends of hexadecanethiol (HC16) and 1H,1H,2H,2H-perfluorooctanethiol (HF6), and NP-C12/F6, coated by mixtures of dodecanethiol (HC12) and HF6; (ii) equal length: NP-C12/F10, coated by HC12 and 1H,1H,2H,2H-perfluorododecanethiol (HF10), and NP-C8/F6, protected by mixtures of octanethiol (HC8) and HF6; and (iii) increased steric hindrance: NP-brC12/F6, featuring mixtures of 3-methyldecan-1-thiol (HbrC12) and HF6. The complete list of the systems investigated in this work is reported in Tables S1–S5, and the synthetic strategies for the preparation of these nanoparticles have been recently described by some of us.
The assessment of the morphology of resulting mixed-monolayer-protected gold NPs may be cumbersome because of a large number of limitations such as their low solubility in common organic solvents, presence of impurities, or the slippery nature of the NP surface, thereby restricting the number of techniques suitable to study their morphology. To overcome some of these practical obstacles, in this work, we adopted an integrated and complementary experimental/computational approach for the characterization of structural features of these mixed SAM-protected gold NPs. Specifically, our experimental analysis relies on 1D and 2D $^{19}$F NMR. NMR techniques based on $^1$H NMR experiments have been successfully used in previous works to characterize monolayer-protected gold nanoparticles and, in particular, to investigate the morphology of mixed monolayers by complexation of lanthanide ions or by NOESY experiments. More recently, an NMR-based approach has been proposed to detail the shell morphology of NPs coated with binary mixtures of aliphatic and aromatic ligands exploiting a combination of chemical shift variations and NOESY experiments. Yet, the $^{19}$F NMR technique is definitely more sensitive to the chemical environment than $^1$H NMR since $^{19}$F chemical shifts span more than 200 ppm, whereas proton chemical shift varies between 0 and 15 ppm. Accordingly, in this work, we exploited this technique further to sensitively probe changes in the environment surrounding the F-ligands as a function of monolayer composition. The theoretical investigation is based on a predictive multiscale molecular simulation protocol, that is, a combination of atomistic/coarse-grained calculations.

**RESULTS AND DISCUSSION**

Gold nanoparticles NP-C16/F6, NP-C8/F6, and NP-brC12/F6 were prepared by direct synthesis, whereas NP-C12/F6 and NP-C12/F10 were obtained by place exchange reaction from narrowly dispersed NP-C12, in turn prepared following the method of Miyake. Synthetic details are reported in ref and, for additional nanoparticles, in the Supporting Information (SI). All samples were fully characterized using UV–vis, transmission electron microscopy (TEM), thermogravimetric analysis, and standard NMR experiments. NP solubility was also assessed and analyzed by increasing the molar fraction of the fluorinated ligands. This property was ultimately rationalized in light of the morphology of the 3D SAM. Monodimensional $^{19}$F NMR experiments were used to monitor the change in chemical shift as a function of the molar fraction of the fluorinated ligands.
a function of the percentage of the fluorinated thiolate in the monolayer. Due to solubility limitations, those nanoparticles soluble only in CHCl₃ were dissolved in CDCl₃ while the remaining samples were solubilized in different mixtures of CDCl₃/C₆F₆, as reported in Tables S1 – S5. All ¹⁹F NMR spectra have been recorded using CFCl₃ as an external reference. The ¹⁹F NMR spectra of the nanoparticles show only two peaks that are well-separated, allowing an accurate measurement of the chemical shift: the peak around −81 ppm represents the CF₃ end group, and the peak at −126 ppm is assigned to the CF₂ group next to the CF₃ (Figure S2). The ¹⁹F NMR spectra indicate that the peaks of nanoparticles shift by changing the composition of the solvent. For this reason, the chemical shifts of the CF₃ and of 7- or 9-CF₂ groups have been corrected for all samples as described in SI for NP-C₁₆/F₆ as an example. Also, the ¹⁹F NMR data reveal that small changes of gold core size, within the same type of NPs, do not influence the chemical shift.

At the same time, multiscale molecular simulations have been carried out on the same set of NPs in order to predict the mixed monolayer organization. Coarse-grained methods as dissipative particle dynamics (DPD) are especially suitable for predicting self-assembling phenomena as those involved in the organization of binary mixtures of ligands on solid curved surfaces because they allow longer sampling times with respect to atomic resolution techniques such as molecular dynamics (MD). The computational approach applied here consists of exploiting information obtained at lower-level simulations (e.g., MD simulations) as input for higher-level calculations (e.g., mesoscopic DPD simulations) in a multiscale framework. DPD simulations were employed to study the effect of mixing different immiscible thiols on the surface of Au NPs in the presence of explicit solvent. With the aim of mimicking the experimental measurement conditions, both CHCl₃ and C₆F₆ were modeled in the relative appropriate amount (Tables S1 – S5). In order to

---

**Figure 4.** Chemical shift (δ) variation of (a) CF₃ groups and (b) 9-CF₂ nuclei increasing the percentage of the fluorinated ligand in the monolayer of NP-C₁₂/F₁₀. Solid line serves as a guide for the eyes only.

**Figure 5.** Equilibrium morphologies of NP-C₁₂/F₁₀ (color code: gray, C₁₂; green, F₁₀) obtained by the mesoscale simulations. Solvent is omitted for clarity.
reproduce a realistic gold core, an icosahedral shape was adopted. Mesoscale models and procedure are described in detail in the SI.

Effect of Solvent Composition on the Chemical Shift.
From the experiments reported in Figures S4 and S5 for NP-C16/F6, it can be observed that the solvent composition has a smaller influence on the chemical shift of the peak corresponding to 7-CF2 groups with respect to that exerted on the CF3 groups. For this reason, we initially surmised that the variation of the chemical shift of CF3 would be larger than that of CF2 groups as a function of the loading of the fluorinated thiolates into the monolayer. On the contrary, even though the CF3 group is more exposed to the solvent than the CF2 group, the variation of their chemical shift as a function of the monolayer composition is similar in both cases. These results further confirm that the chemical shift variations reported here are not a consequence of the variation of the solvent composition but, instead, result from an evolution of the monolayer organization.

Nanoparticles Coated by Branched Ligands, NP-brC12/F6. The chemical shifts of the CF3 and 7-CF2 groups of ligand F6 as a function of the percentage of the fluorinated ligand into the monolayer for the NP-brC12/F6 system are shown in Figure 2. These graphs display that the broad peaks of the nanoparticles gradually shift upfield when the percentage of F6 into the monolayer increases from 0 to 100%. The linear decay of Figure 2 suggests that the average composition of the first nearest neighbor shell of each F-ligand coincides with the overall composition of the monolayer. This is in agreement with the initial hypothesis that the branched structure of the hydrogenated ligand does not allow an ordered crystalline arrangement on the surface of the gold core, hampering the

Figure 6. Chemical shift (δ) variation of (a) CF3 groups and (b) 7-CF2 nuclei increasing the percentage of the fluorinated ligand in the monolayer of NP-C8/F6. Solid line serves as a guide for the eyes only.

Figure 7. Equilibrium morphologies of NP-C8/F6 (color code: gray, C8; green, F6) NPs as obtained by the mesoscale simulations. Solvent is omitted for clarity.
formation of phase-segregated domains. This hypothesis is confirmed by the equilibrium morphologies collected from the corresponding DPD calculations summarized in Figure 3. These simulation structures clearly prove that the two ligands have no tendency to form compact domains, and they prefer to remain isolated; this results in a random organization of the monolayer regardless of the monolayer composition and dimension of the gold core. This evidence agrees well with previous literature data on the organization of monolayers comprising branched thiolates.18

**Nanoparticles Coated by Ligands of Equal Length, NP-C12/F10 and NP-C8/F6.** The behavior of the chemical shifts of the CF$_3$ and 9-CF$_2$ groups versus the percentage of fluorinated ligands into the monolayer for samples of NP-C12/F10 (Figure 4) is different from that observed for NP-brC12/F6, and the interpolation of the points conforms to a 1/(F10%) decay.22 A steep decay is observed when the loading of the fluorinated ligand is less than 40%, indicating a strong evolution of the surface area at the H/F interface; at higher percentages, the chemical shift remains nearly constant, suggesting no significant changes at the H/F interface.

The equilibrium morphologies obtained from the mesoscale simulation of the NP-C12/F10 systems, presented in Figure 5, reveal that the ligands form two distinct domains; that is, the SAM has a Janus-type morphology, in line with a previously reported theoretical prediction on nanoparticles protected by ligands of equal length.16 Indeed, to our knowledge, there are no previous examples of 3D monolayers with Janus morphology obtained by place exchange in the absence of external tools. This organization of the monolayer explains the behavior of the chemical shift of the CF$_3$ and 9-CF$_2$ groups as a function of the monolayer composition.

These results are also supported by the NPs’ tendency to form dimers under TEM conditions, as recently reported for a nanoparticle featuring a Janus organization of the decorating shell.29

---

**Figure 8.** Chemical shift ($\delta$) variation of (a) CF$_3$ groups and (b) 7-CF$_2$ nuclei increasing the percentage of the fluorinated ligand in the monolayer of NP-C16/F6. Solid line serves as a guide for the eyes only.

**Figure 9.** Equilibrium morphologies of selected NP-C16/F6 (color code: gray, C16; green, F6) NPs as obtained by mesoscale simulations. Solvent is omitted for clarity.
At variance with NP-C12/F10, the alternative system featuring ligands of comparable lengths (i.e., NP-C8/F6) is characterized by a different shape of the variation of the chemical shift versus the percentage of F6 in the monolayer. In this case, an intermediate situation between 1/(F6%) and a linear decay is observed (Figure 6). This trend is somewhat unexpected because, intuitively, the monolayers of NP-C8/F6 and the previously discussed NP-C12/F10 should both present a similar Janus-like organization.16 Yet, the corresponding mesoscopic simulations (see Figure 7) indisputably predict that the C8 and F6 thioldates, though similar in length, do not phase-separate in two distinct domains on the surface of the gold core. On the contrary, at small percentages of the fluorinated ligand (i.e., below 20%), the F-thiolates prefer to remain isolated, yielding an essentially random organization. By increasing the loading, the fluorinated ligands do not form extended domains but rather tend to cluster in small, irregular patches. Overall, the morphology of NP-C8/F6 nanoparticles remains ill-defined, as no decisive evidence or reason for a preferred organization can be provided. While deeper and specific investigations on these aspects are underway, a working hypothesis can be anticipated: the shorter chain length of F6 with respect to F10 reduces the overall strength of the fluorophilic interactions, making the formation of fluorinated domains less favorable. In addition, the difference in steric bulk between H- and F-thiolates may represent a bias in the organization of the thiolates when this geometrical mismatch is not counterbalanced by the occurrence of strong fluorophilic interactions.

Nanoparticles Coated by Ligands of Different Length, NP-C16/F6 and NP-C12/F6. The chemical shifts of the CF3 and 7-CF2 groups as a function of the percentage of the F-ligand into the monolayer for the NP-C16/F6 system is shown in Figure 8.

The chemical shift versus composition curve is composed of three regions: an initial, almost linear decay is obtained in the composition range of 0–40%, suggesting that by increasing the

Figure 10. Chemical shift (δ) variation of (a) CF3 groups and (b) 7-CF2 nuclei increasing the percentage of the fluorinated ligand in the monolayer of NP-C12/F6. Solid line serves as a guide for the eyes only.

Figure 11. Equilibrium morphologies of selected NP-C12/F6 (color code: gray, C12; green, F6) NPs as obtained by mesoscale simulations. Solvent is omitted for clarity.
amount of fluorinated ligands into the monolayer these species experience very different chemical surroundings. In the region between 40 and 80%, the chemical shift of the CF₃ group is less sensitive to the composition of the monolayer, indicating that thiolate F₆ experiences very similar chemical environments, even though the percentage of the fluorinated ligand into the monolayer increases. Lastly, when more than 80% of the F₆ thiolate is introduced in the monolayer, the chemical shift of the terminal CF₃ and of 7-CF₂ groups becomes again sensitive to the influence of the neighboring thiolates.

Mesoscale simulations on selected samples of NP-C16/F₆ (Figure 9) revealed that the two ligands organize in striped domains on the surface of the gold NP core, even when 20% of the F₆ thiolate is present in the monolayer. For these NPs, the gain in entropy due to the difference in length and the immiscibility of H- and F-ligands is sufficient to overcome the loss of van der Waals interactions between the hydrogenated alkyl chains, thus leading to a striped structure, in line with previously reported data about the organization of mixed monolayers composed of thiols having different length. Furthermore, the striped organization can explain why some samples of NP-C16/F₆ are soluble in chloroform even at percentages of the fluorinated ligands into the monolayer up to 73%. We reasoned that the sufficiently long hydrogenated chain might mask the short fluorinated ones, screening out the inter-nanoparticle interactions between fluorinated domains if the width of the stripes is small enough, 0.75 nm on average. Evidence from simulations gives further insights into the monolayer organization as a function of the fluorinated thiolate percentage. At values of F₆ less than 40%, increasing the number of fluorinated ligands results in striped domains of F-chains different in size (and, consequently, in interface area), in line with the observed linear decay of the chemical shift. Once the stripes are completely formed (approximately at F₆ > 40%), further addition of fluorinated ligands does not influence the number of thiolates at the H/F interface, as the incoming thiols locate inside a previously formed fluorinated stripe. At percentages of the fluorinated thiolate greater than 80%, F₆ chains prevail, thus reducing the size of the H-domains and modifying the number of thiolates at the interface, ultimately resulting in a substantial decrease of the chemical shift of the diagnostic groups.

For the alternative system characterized by F- and H-thiolates of different length, NP-C12/F₆, the chemical shift of ¹⁹F nuclei in CF₃ and 7-CF₂ as a function of the percentage of the fluorinated ligand is represented by a sigmoidal curve (Figure 10), which is very different from that obtained for the related NP-C16/F₆ system. In this case, a plateau region is found until 25% of fluorinated thiolate is introduced in the monolayer; this trend is followed by a sudden decrease of the chemical shift for small additions of F₆ into the monolayer. Finally, a second plateau region is observed up to 100% F₆. This can be an indication that, at low and high loadings of F₆, there are no marked differences in the surroundings of the fluorinated ligands at the interface triggered by small variations of the monolayer composition. A sensible explanation for this behavior is that, within this interval of compositions, F-/H-regions of comparable size are gradually formed, and this does not influence the overall ratio between thiolates at the H/F interface.

Mesoscale simulations reveal that the monolayer of nanoparticles NP-C12/F₆ does not show a well-defined, ordered organization as a function of the F₆ content. Indeed, as clearly shown in Figure 11, in some instances, stripe-like domains are formed, while at the same time, irregular patches are also observed. Specifically, when 10–15% of the fluorinated ligands are introduced into the monolayer, they form small patches typically composed of a few fluorinated chains. When the loading of F₆ exceeds 30%, elongated patches or stripe-like domains appear, and the dimensions of the relevant domains increase to 0.75–0.80 nm, on average. This is also the composition at which the chemical shift of the CF₃ and 7-CF₂ groups of F₆ starts decreasing, indicating the generation of different environments around the fluorinated ligands. At percentages greater than 60% of F-thiolates, the shape of the domains is again predominantly patchy, and the chemical shift of the CF₃ group becomes insensitive to any further variation of composition, suggesting no important changes in the neighboring environment. The predicted morphologies are also consistent with the solubility properties of NP-C12/F₆. Indeed, with up to 40% of F₆ in the monolayer, these systems are soluble in chloroform, indicating that the surfaces of the fluorinated domains are small enough to be efficiently shielded by the hydrogenated adjacent chains. Increasing the F₆ content leads to an enlargement of the fluorinated domains, and the H-ligands do not hamper interparticle interactions anymore, leading to a change of the solubility properties.

**HOESY Experiments.** NOESY experiments on mixed-monolayer gold nanoparticles enable observation of cross-peaks between the spin systems of two different ligands if their distance is smaller than 0.4 nm and the number of spin systems involved is large enough. These conditions are fulfilled for random and striped mixed monolayers. Similar 2D NMR experiments between F- and H-thiolates, that is, ¹⁹F−¹H HOESY, have been carried out on selected samples of each type of nanoparticle in order to support the morphological analysis described above. In particular, on the basis of the morphologies suggested by 1D NMR experiments and mesoscale simulations, cross-peaks are expected between the CF₃ central groups of the fluorinated ligands and the methylene groups of the hydrogenated ones in the HOESY of nanoparticles NP-brC12/F₆, NP-C16/F₆, NP-C12/F₆, and NP-C8/F₆. Indeed, as reported in Figure S7, the ¹⁹F−¹H HOESY spectra of these nanoparticles with a ratio between F- and H-thiolates close to 1:1 display the presence of cross-peaks between the signal at about −123 ppm, which is assigned to the central CF₃ groups of the F-ligands, and the peak at 1.2 ppm of the methylene groups of the hydrogenated ones. In the HOESY spectra of nanoparticles NP-brC12/F₆ and NP-C16/F₆, a second cross-peak between the same CF₃ central groups and a signal at around 2.5 ppm in the ¹H NMR spectra could also be observed. This cross-peak was assigned to the intramolecular interaction between the CH₂ and CF₂ groups of the same thiolate F₆ as from HOESY experiments on homoligand nanoparticles, NP-F₆, and on the free thiol HF₆.

Unexpectedly, the HOESY spectrum of NP-C12/F10-f presents a cross-peak between the signal of the central CF₂ groups of the fluorinated ligands and the methylene groups of the hydrogenated ones. Since for Janus nanoparticles the fluorinated and hydrogenated ligands are interacting solely at the interface between two domains, the intensity of this cross-peak is expected to be vanishingly small. An explanation for the observation of this cross-peak comes from the length of the fluorinated ligand; thiolate F₁₀ has nine CF₂ groups capable of determining a high number of interactions responsible for the NMR signal. For this reason, even though the F-/H-ligands are interacting exclusively at the interface between the two phase-separated regions, the higher number of CF₂ groups may be sufficient to determine the appearance of the cross-peak. In order to test this hypothesis, we
performed another HOESY analysis on a sample of the same class of nanoparticles presenting a low percentage (13.9%) of the fluorinated thiolate into the monolayer (i.e., the NP-C12/F10-a system). For this system, the fluorinated ligands form a small patch (Figure S), and accordingly, the number of F/H interactions is reasonably lower with respect to NP-C12/F10-f. Yet, even in this case, the HOESY spectrum exhibits a relatively intense cross-peak between the methylene groups of C12 and the central CF2 groups of F10. From these experiments, we conclude that the 19F−1H HOESY is very sensitive and cross-peaks can be observed even though an exiguous number of interactions between the hydrogenated and fluorinated thioclates is present in the monolayer.

CONCLUSIONS

Our analysis of five different set of mixed-monolayer nanoparticles obtained by using blends of hydrogenated and fluorinated ligands and a plethora of different samples for each set nicely covering the loading of the F-ligand over the percentage range offers the basis to develop our understanding of the design rules required to control the formation of specifically patterned monolayer-protected NPs. Specifically, the combination of 19F NMR experiments and mesoscale simulation predictions allows one to assess the morphology of self-organized mixed monolayers comprising mixtures of H- and F-thioclates. Overall, the results point out that, besides the strong immiscibility between H- and F-ligands, other key parameters—such as the high steric hindrance and rigidity of the fluorinated chains—influence the morphology of the organic layer. As a consequence, only when the mismatch of ligand length is equal to eight carbon atoms, stripe-like domains are formed (e.g., the NP-C16/F6 system), driven by entropy gain. On the other hand, a length difference of four carbon atoms gives rise to domains that appear as patches or elongated patches, as seen for the NP-C12/F6 system. Ligands of the same length self-organize in Janus domains (NP-C12/F10), as expected. However, if both ligands are shorter, as in NP-C8/F6, the experimental and theoretical results suggest the formation of a SAM with no specific morphology. Finally, the presence of a branched ligand in the monolayer, such as in the NP-brC12/F6 system, disfavors phase segregation and promotes a random organization of the two types of ligands. The design principles arising from our analysis, although obtained with simple model systems, are envisaged to be of general validity and, as such, are currently being employed by our group in the synthesis and characterization of more complex nanoparticle systems for practical applications.

METHODS

Synthesis. The detailed synthetic procedures for the preparation of gold nanoparticles NP-C16/F6, NP-C12/F6, NP-C12/F10, NP-C8/F6, and NP-brC12/F6 are reported in ref 29. Synthesis of NP-C8/F6-i and NP-C8/F6-p systems is given in full in the SI.

NMR Experiments. NMR experiments were carried out on a Varian 500 spectrometer (operating at 500 MHz for 1H and at 470.08 MHz for 19F). The 1H chemical shifts are referenced to the residual protons in CF2CD2Cl2, whereas CFCl3 was used as an external reference for 19F spectra. The maximum error on 19F chemical shift of repeated measurements is 0.04 ppm.

Samples were prepared by dissolving about 10 mg of NPs in 0.7 mL of deoxygenated CDCl3 or mixtures of CDCl3/C6F6, as detailed in Tables S1−S5 of the SI, and additionally deoxygenated by bubbling argon while keeping the NMR tube in an ultrasound bath for at least 1 min. For 19F−1H HOESY experiments, the pulse sequence FH HOESY was used with 256 scans and a tuning increment, τ1, of 128.

Computational Studies. Coarse-grained methods as DPD are especially suitable for predicting self-assembling phenomena as involved in the organization of binary mixtures of ligands on solid curved surfaces because they allow longer sampling times with respect to atomic resolution techniques (e.g., MD). The computational approach applied here consists of exploiting information obtained at lower-level simulations (e.g., MD simulations) as input for higher-level calculations (e.g., mesoscopic DPD simulations). In detail, a suitable atomistic model of the gold/ligands/solvent interface was employed to estimate the interaction energy among the different system components. These energy values were, in turn, utilized to derive the corresponding DPD simulation parameters. Further, by matching the atomistic/mesoscale pair−pair correlation functions for each ligand, the coarse-grained topology of each chain ligand was calculated. The computational procedure employed here was already successfully applied by our group26,29−31 to predict the self-assembling organization of several immiscible ligand mixtures, including poly(ethylene oxide)-terminating hydrocarbon/perfluorocarbon thiolated chains as well as mercaptoundecanesulfonate/octanethiol ligands on a gold nanoparticle spherical surface and other immiscible polymer-related systems.

ASSOCIATED CONTENT

6 Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsnano.6b03931.

19F NMR of the thiol HF6 and of NP-C12/F6 with peak assignment; description of the experiments to determine the influence of solvent composition on the chemical shift of the NP-C16/F6; preparation of nanoparticles NP-C8/F6-i and NP-C8/F6-p; Tables S1−S5 reporting the characterization data of all nanoparticles, 19F chemical shifts of CF3 and 7- or 9-CF2 groups, and percentage of C6F6 added; 19F−1H HOESY experiments; details of the computational methods, mesoscale models, and mesoscale procedure (PDF)

AUTHOR INFORMATION

Corresponding Authors

*E-mail: paola.posocco@dia.units.it.

*E-mail: l Pasquato@units.it.

Author Contributions

*P. Posocco and L. Pasquato are senior co-authors.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

L. Pasquato and P. Penga dedicate this manuscript to the memory of Professor V. Lucchini, who started the NMR school at the University of Padova and passed away March 22, 2016. The financial support of projects FIRB prot. RBAP11ETKA, MIUR: Project MULTINANOIDA, and University of Trieste (FRA projects 2014 and 2015) is gratefully acknowledged. This work is generously supported by the Italian Ministry of University Research (MIUR) through the Scientific Independence of Young Researchers (SIR) project “Structure and Function at the Nanoparticle biointerface” (Grant RBSI14PBC6 to P.Posocco). The authors are grateful to Prof. Chiara Schmid for her support with the thermogravimetric analyses.

REFERENCES


